Nutrición Hospitalaria

Original

A mixture of *Lactobacillus plantarum* CECT 7315 and CECT 7316 enhances systemic immunity in elderly subjects. A dose-response, double-blind, placebo-controlled, randomized pilot trial

J. Mañé^{1,2*}, E. Pedrosa^{1,2*}, V. Lorén^{1,2}, M. A. Gassull^{1,2}, J. Espadaler³, J. Cuñé³, S. Audivert³, M. A. Bonachera³ and E. Cabré^{1,2,4}

¹Institute for Research in Health Sciences "Germans Trias i Pujol". Badalona. Catalonia. Spain. ²Centro de Investigaciones Biomédicas en Red de Enfermedades Hepáticas y Digestivas (CIBERehd). Barcelona. Spain. ³AB-Biotics. Cerdanyola del Vallès. Catalonia. Spain. ⁴Department of Gastroenterology. Hospital Universitari "Germans Trias i Pujol". Badalona. Catalonia. Spain.

*Contributed equally to the work.

Abstract

Background & aim: Immunosenescence can increase morbi-mortality. Lactic acid producing bacteria may improve immunity and reduce morbidity and mortality in the elderly. We aimed to investigate the effects of a mixture of two new probiotic strains of *Lactobacillus plantarum*—CECT 7315 and 7316— on systemic immunity in elderly.

Methods: 50 institutionalized elderly subjects were randomized, in a double-blind fashion, to receive for 12 weeks 1) 5·10⁸ cfu/day of *L. plantarum* CECT7315/7316 ("low probiotic dose") (n = 13), 2) 5·10⁹ cfu/day of the probiotic mixture ("high probiotic dose") (n = 19), or 3) placebo (n = 15). Leukocyte subpopulations, and cytokine levels (IL-1, IL-10, TGF- β 1) were measured in venous blood at baseline, end of treatment (week 12), and end of follow-up (week 24). Infection and survival rates were recorded.

Results: After treatment, high probiotic dose resulted in significant increases in the percentages of activated potentially T-suppressor (CD8+CD25+) and NK (CD56+CD16+) cells, while low probiotic dose increased activated T-helper lymphocytes (CD4+CD25+), B lymphocytes (CD19+), and antigen presenting cells (HLA-DR+). Also, plasma TGF- β 1 concentration significantly decreased after treatment with both probiotic doses. Most of these changes remained 12 weeks after probiotic discontinuation. Incidence of infections during treatment showed a significant trend to be lower in the high probiotic dose group. In addition, there was a significant trend for mor-

Correspondence: Eduard Cabré. Department of Gastroenterology. Hospital Universitari Germans Trias i Pujol. Carretera del Canyet, s/n. 08916 Badalona. Spain. E-mail: ecabre.germanstrias@gencat.cat

Recibido: 1-XI-2010. Aceptado: 5-XI-2010.

UNA MEZCLA DE *LACTOBACILLUS PLANTARUM* CECT 7315 Y CECT 7316 MEJORA LA INMUNIDAD SISTÉMICA EN ANCIANOS. UN ENSAYO ALEATORIO PILOTO, DE DOSIS-RESPUESTA, DOBLE CIEGO Y CONTROLADO CON PLACEBO

Resumen

Introducción y objetivos: La inmunosenescencia puede aumentar la morbi-mortalidad. Las bacterias productoras de ácido láctico pueden mejorar la inmunidad y disminuir la morbilidad y mortalidad en los ancianos. Nuestro objetivo fue investigar los efectos de una mezcla de dos nuevas cepas probióticas de *Lactobacillus plantarum* —CECT 7315 y 7316— sobre la inmunidad sistémica en ancianos.

Métodos: 50 ancianos institucionalizados se aleatorizaron, en un diseño a doble-ciego, para recibir durante 12 semanas 1) 5·10⁸ ufc/día de *L. plantarum* CECT7315/7316 ("dosis baja de probiótico") (n = 13), 2) 5·10⁹ ufc/día de la mezcla probiótica ("dosis alta de probiótico") (n = 19), o 3) placebo (n = 15). Se determinaron las subpoblaciones leucocitarias y los niveles de citokinas (IL-1, IL-10, TGF-β1) en sangre venosa periférica basalmente, al final del tratamiento (sem. 12) y al final del seguimiento (sem. 24). Se registró la tasa de infecciones y la mortalidad.

Resultados: Tras el tratamiento, la dosis alta de probiótico indujo aumentos significativos en los porcentajes de células potencialmente T-supresoras (CD8+CD25+) y NK (CD56+CD16+) activadas, en tanto que la dosis baja aumento los linfocitos T-colaboradores activados (CD4+CD25+), los linfocitos B (CD19+), y las células presentadoras de antígeno (HLA-DR+). Asimismo, la concentración plasmática de TGF- β 1 disminuyó tras el tratamiento con ambas dosis de probiótico. La mayor parte de estos cambios se mantuvieron 12 semanas después de suspender el tratamiento. La incidencia de infecciones durante el tratamiento mostró una tendencia significativa a ser menor con la dosis alta de probiótico, mientras que se observó una tendencia significativa a que la mortalidad tality to be greater in the placebo group *vs.* both probiotic groups.

Conclusions: Depending on the dose, *L. plantarum* CECT7315/7316 have different immune-enhancing effects in elderly subjects. These effects might result in a better clinical outcome.

(Nutr Hosp. 2011;26:228-235)

DOI:10.3305/nh.2011.26.1.5112

Key words: *Immune cells*. *Cytokines*. *Aging*. *Probiotics*. Lactobacillus plantarum.

Introduction

Some lactic acid bacteria strains are defined as probiotic as far as they are able to confer a variety of physiologic benefits to the host.¹ Dietary supplementation with certain *Bifidobacteria* and *Lactobacilli* strains has been proven to be useful in the management of several gastrointestinal disorders such as acute infectious and antibiotic associated diarrhea in children^{2,3} and adults,⁴ ulcerative colitis,⁵⁻⁹ pouchitis,^{10,11} and irritable bowel syndrome.^{12,13} Also, they have been reported to be useful in protecting children from allergic illnesses,^{14,15} and in the prevention of bacterial, fungal or viral infections.^{16,17}

Human studies have revealed that probiotic bacteria can have an influence on the host's immune system. Some components of the immune response, including phagocytosis, natural killer (NK) cell activity and mucosal IgA production (especially in children), can be improved by some probiotic bacteria.^{18,19} Other components, including lymphocyte proliferation, the production of cytokines and antibodies other than IgA appear less sensitive to probiotics.^{18,19}

It is well characterized that aging involves an involution and decreases the capacity to mediate effective immune responses to vaccination and invading pathogens.^{20,21} Immunosenescence has been associated to a decrease of mature T cell numbers, changes of NK and dendritic cell proportions, and loss of diversity of B cells in the blood of the elderly.^{20,22} Moreover, aging causes a decline in cell-mediated cytotoxic and phagocytic responses, and increases circulating levels of proinflammatory cytokines.²⁰ Clinically, these changes potentially increase morbidity and mortality in elderly individuals through an increased rate of infections, malignancy, and autoimmunity-related disorders.²¹

Intervention trials in elderly subjects have shown that oral supplementation with *Bifidobacterium lactis* HN019 significantly increases the proportion of total, helper (CD4+) and activated (CD25+) lymphocytes in peripheral blood.²³ In addition, *Lactobacillus rhamnosus* HN001 enhanced the *ex vivo* phagocytic capacity of polymorphonuclear leukocytes, as well as the tumoricidal activity of NK cells in these individuals.²⁴ Furthermore, it has been shown that elderly people receivfuera mayor el grupo placebo vs. ambos grupos tratados con probiótico.

Conclusiones: Dependiendo de la dosis, *L. plantarum* CECT7315/7316 tiene distintos efectos inmunoestimulantes en ancianos. Dichos efectos podrían contribuir a una mejor evolución clínica.

(*Nutr Hosp*. 2011;26:228-235)

DOI:10.3305/nh.2011.26.1.5112

Palabras clave: Inmunocitos. Citokinas. Envejecimiento. Probióticos. Lactobacillus plantarum.

ing fermented milk with *Lactobacillus casei* DN-114001 reduce the duration of upper respiratory tract infections.^{25,26} Therefore, functional foods containing probiotics might have a particular applicability in elderly population.

The present study primarily aimed to investigate the effects on systemic immunity of a mixture of two new probiotic strains of *Lactobacillus plantarum* —CECT 7315 and CECT 7316— at two different dosages, in institutionalized elderly subjects by means of a randomized, double-blind, placebo-controlled pilot trial. Clinical outcomes including infection and survival rates were also recorded.

Subjects and methods

Probiotic strains

Two new strains of *L. plantarum* showing probiotic properties²⁷ (CARINSA, Barcelona, Spain) were used in this trial. They were originally isolated from the feces of an infant, and identified in the Spanish Collection of Type Cultures (CECT) as CECT 7315 and CECT 7316. Both strains showed a high survival rate both in acidic conditions and in presence of bile acids *in vitro*.²⁷ Also, they showed a high capacity of adherence to pig intestinal cells in culture, doubling those of *L. reuteri* and *L. rhamnosus GG*, and their pathogen antagonism is quite similar to that of *L. rhamnosus GG*²⁷. For the purposes of the present trial both strains were mixed at a 1:1 ratio.

Subjects and trial design

This trial was conducted, in winter 2006 and spring 2007, in subjects older than 65, institutionalized in two geriatric centers in the metropolitan area of Barcelona, Spain: "Llars Mundet" (Barcelona, Spain) and "Centre Assistencial Dr. Emili Mira" (Sta. Coloma de Gramenet, Spain). Exclusion criteria were 1) suffering from any acute illness during the previous month, 2) having a neoplastic disease, 3) a life expectancy lower than 6 months, 4) documented intolerance to milk or dairy

products, 5) swallowing disturbances, and 6) use of antibiotics, probiotics, nutritional supplements and/or functional foods in the previous month.

Subjects were randomized to receive, in a doubleblind fashion, either 1) $5 \cdot 10^8$ cfu/day of the *L. plantarum* CECT7315/7316 mixture in 20 g of powdered skimmed milk ("low probiotic dose"), 2) $5 \cdot 10^9$ cfu/day of the probiotic mixture in the same vehicle ("high probiotic dose"), or 3) vehicle alone (placebo). Randomization was performed in separate lists for each participating center. Both probiotic preparations and placebo were presented in identical vacuum sealed sachets, and administered as single daily dose, diluted in 200 mL of water or other cold drink, for 12 weeks. Subjects were then followed for a period of 12 additional weeks (follow-up period).

Medical visits were performed at baseline, and every 4 weeks until the end of the follow-up period (week 24). Clinical anamnesis (with particular reference to infections) and physical examination were carried out at every visit. Infection was defined as either 1) a febrile episode with a definite clinical focus and/or microbiologically confirmed etiology, or 2) a febrile episode without clinical focus or positive culture, but requiring empirical antibiotic therapy.

The body mass index (BMI), and the Barthel index to assess the functional capacity for daily living activities,²⁸ were measured at baseline, and weeks 12 (end of the treatment period) and 24 (end of follow-up). Also, fasting venous blood samples were obtained at these time points for routine laboratory analysis, and immunological parameters evaluation (see below).

Blood immunological parameters

The effect of the probiotic mixture on blood immunological parameters was the primary end point of this trial. As mentioned, they were measured at baseline, at the end of the treatment period (week 12), and at the end of the follow-up period (week 24).

Assessment of blood leukocyte subpopulations

Leukocytes were obtained from heparinized whole blood samples by means of Lymphoprep[™] (Axis-Shield, Oslo, Norway) gradient centrifugation. After being washed in cool PBS, freshly isolated leukocytes were incubated for 15 min. in darkness with fluorochrome-conjugated mouse antibodies in two separate tubes. In the first one, different subsets of T cells, active or not, were labeled with anti-CD3-PECy5, anti-CD4-FITC, anti-CD8-APC and anti-CD25-PE (BD Biosciences, San José, CA, USA). In the second tube, total T cells, NK cells, B cells and antigen-presenting cells were stained by anti-CD3-FITC, anti-CD56/16-PE, anti-CD19-APC, and anti-HLA-DR-PECy5 (BD Biosciences, San José, CA, USA), respectively. Phenotypic assessment was performed by flow cytometry using a FACScalibur cytometer (Becton Dickinson, Frankin Lakes, NJ, USA).

Plasma cytokine measurements

Plasma concentrations of IL-1 and IL-10 were measured by BDTM Cytometric Beat Array Human Soluble Proteins Flex Set assays (BD Biosciences, San Diego, USA) according to the manufacturer's protocols. The beat analyses were resolved in FL3 and FL4 channels of the BD FACScalibur flow cytometer (BD Biosciences, San José, CA, USA). Analyses of sample data were performed using FCAP Array[™] software (BD Biosciences, San José, CA, USA).

Plasma concentrations of TGF- β 1 were evaluated using a commercially available ELISA kit (Deltaclon, Madrid, Spain) following the manufacturer's instructions. Plasma samples were acidified with HCl 1 M, and diluted to 1:50 prior to analysis. All measurements were performed in duplicated.

Ethical considerations

All subjects gave their informed consent to participate in the study before randomization. The trial was performed under the norms of the Helsinki's Declaration, and was approved by the Ethical Committees of the Institute for Research in Biomedical Sciences Germans Trias i Pujol, and the participating centers.

Statistical analysis

Data are expressed as median plus interquartile range (IQR) for quantitative parameters, and as frequencies for qualitative variables.

Comparisons of quantitative variables among groups were performed by means of the Kruskal-Wallis non-parametric test (with post-hoc Mann-Whitney U test). Quantitative variables among groups were compared with the Chi² test. Changes in quantitative variables within groups were assessed by means of the Wilcoxon rank-sum test for repeated measures. All statistical analyses were performed using the SPSS 12.0 package for Windows (SPSS, Chicago, IL, USA). Pvalues below 0.05 were considered as significant.

Results

A total of 60 individuals were assessed for eligibility and randomized (n = 20 for each therapeutic group). Unfortunately, however, 10 subjects withdrew their consent within the first 72 hours after randomization. Thus, 50 subjects were finally included in the study (fig 1).²⁹ There were no differences among the therapeutic



groups at baseline, regarding demographics, BMI, Barthel index, and routine laboratory parameters (table I). No adverse events attributable to the trial supplements were recorded.

Immunological parameters

Immunological parameters —the main end-point of the study— could be only assessed in those individuals surviving the treatment period (15 in the placebo group, 13 in the low-dose probiotic group, and 19 in the high-dose probiotic group) (fig. 1).

Blood leukocyte subpopulations

Baseline percent values of the different cell phenotypes were similar among the three therapeutic groups (table II). *L. plantarum* CECT 7315/7316 induced different changes in blood leukocyte subpopulations depending on the dose of probiotic administered. At the end of the treatment (week 12) high dose resulted in significant increases in the percentages of activated potentially T-suppressor (CD8+CD25+) and NK (CD56+CD16+) cells, while low dose induced increases in activated T-helper lymphocytes (CD4+ CD25+), B lymphocytes (CD19+), and antigen presenting cells (HLA-DR+) (table II). Of note, most of these changes remained at the end of the follow-up period (week 24), 12 weeks after probiotic treatment cessation (table II).

Plasma cytokine concentrations

Plasma concentrations of both IL-1 and IL-10 were undetectable at every time point in all therapeutic groups. Plasma TGF- β 1 levels were similar at baseline among the three groups. A significant decrease in TGF- β 1 concentration was observed after treatment with both probiotic doses, and at the end of follow-up period, while no change was observed in the placebo group (fig. 2).

Clinical outcomes

Seven subjects of the placebo group developed infections: 4 of them during the 12-week therapeutic period (3 fatal cases of pneumonia, one case of urinary tract infection), and 3 during the follow-up period (acute bronchitis in 2 cases, urinary tract infection in one case). Five individuals of the group treated with low probiotic suffered from infections: 3 during the treatment period (pneumonia in 2 cases, acute bronchitis in one) and 2 during the follow-up (acute bronchitis, middle otitis). Three subjects in the high probiotic dose suffered from infections during the follow-up (acute bronchitis in 2, urinary tract infection in one). The

Table IBaseline characteristics of the three therapeutic groups						
	Placebo (n = 18)	Low-dose probiotic $(n = 13)$	High-dose probiotic $(n = 19)$	p^*		
Age (years)*	69 (66-82)	70 (67-83)	71 (65-84)	NS		
Male gender (n)	10	6	10	NS		
BMI (Kg/m ²)*	26.9 (25.8-28.9)	24.4 (23.0-27.8)	25.7 (24.2-29.0)	NS		
Barthel index (I/MD/PD/SD/FD)1	1/13/3/1/0	3/9/1/0/0	0/16/1/1/1	NS		
s-Albumin (g/L)*	37 (36-39)	38 (35-39)	38 (34-39)	NS		
s-Glucose (mg/dL)*	105.3 (95.3-117.0)	100.9 (97.2-115.1)	107.0 (99.3-125.0)	NS		
s-Total Cholesterol (mg/dL)*	206.1 (199.0-236.1)	211.1 (203.1-243.3)	205.0 (202.3-250.0)	NS		
s-Triglycerides (mg/dL)*	128.6 (123.1-178.9)	140.7 (125.3-188.1)	134.8 (130.2-200.3)	NS		
s-Creatinine (mg/dL)*	0.92 (0.80-1.30)	0.95 (0.91-1.17)	0.91 (0.78-1.35)	NS		
s-AST (IU/L)*	27 (26-32)	30 (25-37)	28 (26-39)	NS		
s-ALT (IU/L)*	32 (28-41)	31 (29-39)	30 (26-40)	NS		
s-Alk. Phosphatase (IU/L)*	113 (97-129)	100 (86-128)	101 (95-154)	NS		
s-GGT (IU/L)*	29 (26-46)	34 (25-50)	32 (26-64)	NS		
s-Total Bilirubin (mg/dL)*	0.67 (0.45-1.10)	0.55 (0.60-0.99)	0.50 (0.40-1.23)	NS		
Hemoglobin (g/dL)*	13.3 (12.5-15.1)	14.1 (12.0-14.9)	13.6 (11.9-15.7)	NS		
Leukocytes (x10 ⁹ /L)*	8.21 (7.65-9.93)	7.95 (6.45-10.1)	8.11 (7.50-9.89)	NS		
Platelets (x10 ⁹ /L)*	222 (201-340)	205 (199-400)	216 (202-417)	NS		

*Kruskal-Wallis test. *median (IOR)

Independent/Mildly dependent/Partly dependent/Severely dependent/Fully dependent.



Fig. 2.—Time-course of TGF- $\beta 1$ concentration in the three groups. "p < 0.05 vs. baseline; "p < 0.01 vs. baseline (Wilcoxon rank-sum test).

infection rate during the treatment period showed a significant trend to be lower in the high probiotic dose group, while there were no differences in the follow-up period (table III).

As mentioned, the 3 cases of pneumonia in the placebo group were the only deaths occurring in the study. Thus, there was a significant trend for mortality to be greater in the placebo group as compared to the probiotic groups (table III). No case of mortality occurred during the follow-up.

No significant change in the BMI, the Barthel index, and the routine laboratory test was observed in the survivors of the three groups, either during the treatment or the follow-up periods (data not shown).

Discussion

In the last two decades, growing evidence has been produced stressing the role of the intestinal microbiota in the development of both local and systemic immunity.^{30,31} This regulatory activity involves intestinal epithelial cells, macrophages, dendritic cells, and Tlymphocytes. In fact, axenic mice have been shown to posses fewer dendritic cells in the gut-associated lymphoid tissue, smaller amounts of T-cells in the spleen, and decreased activation of CD4+ cells than animals with normal intestinal microbiota.^{32,34}

Only a few of the vast number of *L. plantarum* strains identified to date have well established immunomodulatory properties. *L. plantarum* CECT 7315 and 7316 strains were identified as probiotics as a result of extensive studies on different bacterial strains isolated from 0-5 year-old children mostly fed with vegetables.²⁷ The results of the present study with (institutionalized) healthy elders show that supplementation with the *L. plantarum* CECT 7315/7316 combination is effective in enhancing systemic immunity in humans, resulting in increased numbers of B lymphocytes (CD19+), NK (CD56+CD16+) cells, and antigen presenting cells (HLA-DR+), in addition to enhanced activation (CD25 expression) of CD4+ and CD8+ T-

%		Baseline [#]	End of treatment (week 12)	End follow-up (week 24)
CD3+	Placebo	67.8 (51.9-73.5)	61.0 (59.6-71.8)	64.3 (56.2-69.0)
	Low-dose probiotic	71.1 (56.5-73.1)	67.7 (56.6-75.6)	64.7 (58.6-73.3)
	High-dose probiotic	66.8 (59.1-75.6)	71.6 (58.0-76.4)	66.5 (59.5-72.1)
CD4+	Placebo	37.7 (32.3-45.0)	34.1 (27.8-43.3)	37.5 (32.6-40.4)
	Low-dose probiotic	37.5 (27.4-46.8)	36.1 (30.1-44.9)	36.1 (30.8-45.2)
	High-dose probiotic	35.9 (24.9-42.8)	30.9 (26.3-38.9)	30.0 (24.9-38.4)
CD4+CD25+	Placebo	13.9 (10.3-22.4)	14.2 (13.3-26.4)	15.3 (12.4-22.3)
	Low-dose probiotic	12.2 (8.9-18.0)	17.0 (11.1-22.2)*	14.3 (10.2-21.9)
	High-dose probiotic	11.6 (10.6-20.2)	15.4 (11.0-21.3)	14.5 (9.8-20.1)
CD8+	Placebo	21.3 (12.2-26.3)	18.5 (12.9-26.2)	21.7 (13.2-28.4)
	Low-dose probiotic	20.6 (13.3-28.4)	19.1 (12.3-30.4)	19.4 (12.9-30.1)
	High-dose probiotic	22.2 (17.7-36.4)	23.4 (21.7-36.9)	23.5 (20.6-37.7)
CD8+CD25+	Placebo	3.3 (1.6-4.8)	3.2 (2.2-4.5)	3.1 (2.5-4.1)
	Low-dose probiotic	3.4 (2.2-4.9)	3.7 (2.6-6.3)	3.9 (2.3-4.9)
	High-dose probiotic	2.9 (2.1-4.9)	3.8 (2.9-6.0)*	3.9 (2.4-6.1)*
CD19+	Placebo	6.3 (6.0-8.3)	6.4 (6.2-10.7)	6.4 (5.9-10.4)
	Low-dose probiotic	5.9 (5.1-8.4)	6.8 (6.2-9.1)*	7.6 (5.4-11.9)*
	High-dose probiotic	6.7 (4.9-7.5)	6.8 (4.8-7.9)	6.7 (4.5-7.8)
CD56+CD16+	Placebo	18.1 (10.3-28.6)	18.4 (14.3-27.1)	18.4 (15.5-26.9)
	Low-dose probiotic	17.2 (11.4-25.5)	19.5 (11.4-27.9)	17.2 (14.4-26.9)
	High-dose probiotic	16.9 (10.4-24.4)	19.8 (10.4-32.8)*	18.0 (12.5-33.5)*
HLA-DR+	Placebo	6.8 (6.3-8.3)	6.1 (5.8-7.4)	7.3 (5.9-8.6)
	Low-dose probiotic	6.5 (6.2-9.3)	7.5 (7.2-9.2)*	7.5 (6.4-9.7)*
	High-dose probiotic	6.3 (5.6-8.1)	6.3 (5.6-7.0)	6.0 (5.0-6.9)

Table II

*No differences among the three groups were observed at baseline for none of the phenotypes (Kruskal-Wallis test).

*P < 0.03 vs. Baseline (Wilcoxon rank-sum test)*Independent/Mildly dependent/Partly dependent/Severely dependent/Fully dependent.

cells. These results are in agreement with previous studies which have shown that the intake of lactic acid producing bacteria increases the CD4+, CD25+, CD19+ and CD56+ phenotypes in peripheral blood cells from elderly volunteers.^{23,35} Taken together, these findings prove the usefulness of probiotic bacteria to cope with the process of immunosenescence in the elderly.

One of the aims of the study was to assess which of two probiotic dosages had the greatest immune enhancing effects. Unexpectedly, however, we found not quantitative but qualitative differences in the immunological effects between the two evaluated dosages. Supplementation with 5·10⁸ cfu/day of the *L. plantarum* CECT7315/7316 mixture (low dose) was associated with changes in the blood cell subsets suggesting an enhanced immuneregulatory and/or Th2 polarized response (increased CD4+CD25+, CD19+, and HLA-DR+ cells).³⁶⁻³⁸ In contrast, using a daily probiotic dose ten times greater (high dose) resulted in a significant increase of potentially cytotoxic (CD8+ CD25+, CD56+CD16+) cell phenotypes³⁹ in peripheral blood.

These findings open the possibility to use different probiotic dosage for different indications. For instance,

one could anticipate that low doses might be useful as coadjuvant therapy to vaccinations^{40,41} —as far as they promote acquired humoral immune responses—, while higher doses might be useful to prevent infections¹⁷ – as they promote more immediate and unspecific cellular responses.

The observed decrease in plasma TGF-β1 concentration associated to *L. plantarum* CECT 7315/7316 supplementation, no matter which dose was administered, deserves special comment. TGF-β1 belongs to a superfamily of cytokines which regulate a plethora of developmental processes, and a disruption of their activity has been involved in a variety of human diseases ranging from fibrotic diseases to the progression of many cancers.⁴² Immunological actions of TGF-β1 include inhibition of dendritic cell maturation and NK activity.⁴³ More recently, its fundamental role in the polarization of the Th17 response has been related to highly pro-inflammatory T cell subset and to some autoimmune processes,^{44,45} which could be particularly relevant in the elderly.⁴⁶

All these immunomodulatory actions of *L. plantarum* CECT 7315/7316 might have a positive impact on clinical outcome. Indeed, in spite that the present trial is clearly underpowered to assess clinical end points, a

Table III
Infactions and montality during the treatment and follow up periods in the three therapeutic group

ingections and morial	ngections and mortality during the realment and jonow-up periods in the infect merupeance groups					
	Placebo	Low-dose probiotic	High-dose probiotic	p for trend*		
Infections during treatment period	4/18	3/13	0/19	0.049		
Infections during follow-up period	3/15	2/13	3/19	NS		
Mortality during treatment period	3/18	0/13	0/19	0.037		
Mortality during follow-up period	0/15	0/13	0/19	NS		

*Chi-square test.

positive effect of probiotic supplementation on both infection rate and survival is suggested. Nevertheless, these promising data must be confirmed in larger trials specifically designed to assess clinical outcomes.

Conflict of interest statement

JM, MAG, JE, JC, SA, MAB, and EC share the authorship of the patent involving the probiotic strains used in this trial. JE, JC, SA, and MAB are affiliates of AB-BIOTICS, the company that developed the probiotic strains.

Statement of authorship

JM and EP contributed to the design of the trial, immunological measurements and drafting of manuscript. VL performed the immunological studies. MAG contributed to the design of the trial. JE, SA, and MAB contributed to the design of the trial and provided significant advice on the properties of the probiotic strains used. JC collected clinical data and performed the statistical analysis. EC contributed to the design of the study, collection of data, statistical analysis and drafting of the manuscript. The final version of the manuscript has been approved by all the authors.

Acknowledgements

The authors are in debt with Dr. Margarita Méndez-Sánchez (from "Llars Mundet", Barcelona, Spain), and Dr. Montserrat Pérez-Carre (from "Center Assistencial Dr. Emili Mira", Sta. Coloma de Gramenet, Spain).

This study was funded by CARINSA, ACC10 (Catalan Government) and PROFIT-CDTI (Spanish Government).

References

- Heczko PB, Strus M, Kochan P. Critical evaluation of probiotic activity of lactic acid bacteria and their effects. *J Physiol Phar*macol 2006; 57 (Suppl. 9): 5-12.
- 2. Szajewska H, Mrukowicz JZ. Probiotics in the treatment and prevention of acute infectious diarrhea in infants and children: a

systematic review of published randomized, double-blind, placebo-controlled trials. *J Pediatr Gastroenterol Nutr* 2001; 33 (Suppl. 2): S17-S25.

- Van Niel CW, Feudtner C, Garrison MM, Christakis DA. Lactobacillus therapy for acute infectious diarrhea in children: a meta-analysis. *Pediatrics* 2002; 109: 678-684.
- 4. Gao XW, Mubasher M, Fang CY, Reifer C, Miller LE. Doseresponse efficacy of a proprietary probiotic formula of Lactobacillus acidophilus CL1285 and Lactobacillus casei LBC80R for antibiotic-associated diarrhea and Clostridium difficileassociated diarrhea prophylaxis in adult patients. *Am J Gastroenterol* 2010; 105: 1636-1641.
- Ishikawa H, Akedo I, Umesaki Y, Tanaka R, Imaoka A, Otani T. Randomized controlled trial of the effect of bifidobacteriafermented milk on ulcerative colitis. *J Am Coll Nutr* 2002; 22: 56-63.
- 6. Kato K, Mizuno S, Umesaki Y, Ishii Y, Sugitani M, Imaoka A, et al. Randomized placebo-controlled trial assessing the effect of bifidobacteria-fermented milk on active ulcerative colitis. *Aliment Pharmacol Ther* 2004; 20: 1133-1141.
- Tursi A, Brandimarte G, Giorgetti GM, Forti G, Modeo ME, Gigliobianco A. Low-dose balsalazide plus a high-potency probiotic preparation is more effective than balsalazide alone or mesalazine in the treatment of acute mild-to-moderate ulcerative colitis. *Med Sci Monit* 2004; 10: 126-131.
- 8. Miele E, Pascarella F, Giannetti E, Quaglietta L, Baldassano RN, Staiano A. Effect of a probiotic preparation (VSL#3) on induction and maintenance of remission in children with ulcerative colitis. *Am J Gastroenterol* 2009; 104: 437-443.
- Sood A, Midha V, Makharia GK, Ahuja V, Singal D, Goswami P, et al. The probiotic preparation, VSL#3 induces remission in patients with mild-to-moderately active ulcerative colitis. *Clin Gastroenterol Hepatol* 2009; 7: 1202-9, 1209.
- 10. Elahi B, Nikfar S, Derakhshani S, Vafaie M, Abdollahi M. On the benefit of probiotics in the management of pouchitis in patients underwent ileal pouch anal anastomosis: a meta-analysis of controlled clinical trials. *Dig Dis Sci* 2008; 53: 1278-1284.
- 11. Gionchetti P, Rizzello F, Morselli C, Poggioli G, Tambasco R, Calabrese C et al. High-dose probiotics for the treatment of active pouchitis. *Dis Colon Rectum* 2007; 50: 2075-2082.
- 12. Moayyedi P, Ford AC, Talley NJ, Cremonini F, Foxx-Orenstein A, Brandt LJ et al. The efficacy of probiotics in the therapy of irritable bowel syndrome: a systematic review. *Gut* 2010; 59: 325-332.
- 13. Brenner DM, Moeller MJ, Chey WD, Schoenfeld PS. The utility of probiotics in the treatment of irritable bowel syndrome: a systematic review. *Am J Gastroenterol* 2009; 104: 1033-1049.
- 14. Osborn DA, Sinn JK. Probiotics in infants for prevention of allergic disease and food hypersensitivity. *Cochrane Database Syst Rev* 2007; CD006475.
- Kalliomaki M, Antoine JM, Herz U, Rijkers GT, Wells JM, Mercenier A. Guidance for substantiating the evidence for beneficial effects of probiotics: prevention and management of allergic diseases by probiotics. *J Nutr* 2010; 140: 713S-721S.
- De Vrese M., Winkler P, Rautenberg P, Harder T, Noah C, Laue C et al. Effect of Lactobacillus gasseri PA 16/8, Bifidobacterium longum SP 07/3, B. bifidum MF 20/5 on common

cold episodes: a double blind, randomized, controlled trial. *Clin Nutr* 2005; 24: 481-491.

- 17. Wolvers D, Antoine JM, Myllyluoma E, Schrezenmeir J, Szajewska H, Rijkers GT. Guidance for substantiating the evidence for beneficial effects of probiotics: prevention and management of infections by probiotics. *J Nutr* 2010; 140: 698S-712S.
- Nova E, Warnberg J, Gomez-Martinez S, Diaz LE, Romeo J, Marcos A. Immunomodulatory effects of probiotics in different stages of life. *Br J Nutr* 2007; 98 (Suppl. 1): S90-S95.
- Lomax AR, Calder PC. Probiotics, immune function, infection and inflammation: a review of the evidence from studies conducted in humans. *Curr Pharm Des* 2009; 15: 1428-1518.
- Panda A, Arjona A, Sapey E, Bai F, Fikrig E, Montgomery RR et al. Human innate immunosenescence: causes and consequences for immunity in old age. *Trends Immunol* 2009; 30: 325-333.
- 21. Agarwal S, Busse PJ. Innate and adaptive immunosenescence. Ann Allergy Asthma Immunol 2010; 104: 183-190.
- Gibson KL, Wu YC, Barnett Y, Duggan O, Vaughan R, Kondeatis E et al. B-cell diversity decreases in old age and is correlated with poor health status. *Aging Cell* 2009; 8: 18-25.
- Gill HS, Rutherfurd KJ, Cross ML, Gopal PK. Enhancement of immunity in the elderly by dietary supplementation with the probiotic Bifidobacterium lactis HN019. *Am J Clin Nutr* 2001; 74: 833-839.
- Sheih YH, Chiang BL, Wang LH, Liao CK, Gill HS. Systemic immunity-enhancing effects in healthy subjects following dietary consumption of the lactic acid bacterium Lactobacillus rhamnosus HN001. J Am Coll Nutr 2001; 20: 149-156.
- Turchet P, Laurenzano M, Auboiron S, Antoine JM. Effect of fermented milk containing the probiotic Lactobacillus casei DN-114001 on winter infections in free-living elderly subjects: a randomised, controlled pilot study. *J Nutr Health Aging* 2003; 7: 75-77.
- Guillemard E, Tondu F, Lacoin F, Schrezenmeir J. Consumption of a fermented dairy product containing the probiotic Lactobacillus casei DN-114001 reduces the duration of respiratory infections in the elderly in a randomised controlled trial. *Br J Nutr* 2010; 103: 58-68.
- Martinez V, López Q, Gassull MA, Cuñé J, Espadaler J, Cabré E, et al. Strains of *Lactobacillus plantarum* as probiotics. *European Patent* 2007; 07121817.6.
- 28. Mahoney FI, Barthel DW. Functional evaluation: The Barthel index. *Md State Med J* 1965; 14: 61-65.
- Moher D, Schulz KF, Altman DG. The CONSORT statement: revised recommendations for improving the quality of reports of parallel-group randomized trials. *Ann Intern Med* 2001; 134: 657-662.
- Mazmanian SK, Kasper DL. The love-hate relationship between bacterial polysaccharides and the host immune system. *Nat Rev Immunol* 2006; 6: 849-858.
- Round JL, Mazmanian SK. The gut microbiota shapes intestinal immune responses during health and disease. *Nat Rev Immunol* 2009; 9: 313-323.

- Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell* 2005; 122: 107-118.
- Walton KL, He J, Kelsall BL, Sartor RB, Fisher NC. Dendritic cells in germ-free and specific pathogen-free mice have similar phenotypes and in vitro antigen presenting function. *Immunol Lett* 2006; 102: 16-24.
- Ostman S, Rask C, Wold AE, Hultkrantz S, Telemo E. Impaired regulatory T cell function in germ-free mice. *Eur J Immunol* 2006; 36: 2336-2346.
- Gill HS, Rutherfurd KJ, Cross ML. Dietary probiotic supplementation enhances natural killer cell activity in the elderly: an investigation of age-related immunological changes. J Clin Immunol 2001; 21: 264-271.
- 36. Sakaguchi S, Sakaguchi N, Shimizu J, Yamazaki S, Sakihama T, Itoh M, et al. Immunologic tolerance maintained by CD25+ CD4+ regulatory T cells: their common role in controlling autoimmunity, tumor immunity, and transplantation tolerance. *Immunol Rev* 2001; 182: 18-32.
- Mizoguchi A, Bhan AK. A case for regulatory B cells. J Immunol 2006; 176: 705-710.
- Bulwin GC, Walter S, Schlawinsky M, Heinemann T, Schulze A, Hohne W et al. HLA-DR alpha 2 mediates negative signalling via binding to Tirc7 leading to anti-inflammatory and apoptotic effects in lymphocytes in vitro and in vivo. *PLoS One* 2008; 3:e1576.
- Zamai L, Ahmad M, Bennett IM, Azzoni L, Alnemri ES, Perussia B. Natural killer (NK) cell-mediated cytotoxicity: differential use of TRAIL and Fas ligand by immature and mature primary human NK cells. *J Exp Med* 1998; 188: 2375-2380.
- Vintini E, Villena J, Alvarez S, Medina M. Administration of a probiotic associated with nasal vaccination with inactivated Lactococcus lactis-PppA induces effective protection against pneumoccocal infection in young mice. *Clin Exp Immunol* 2010; 159: 351-362.
- Soh SE, Ong DQ, Gerez I, Zhang X, Chollate P, Shek LP, et al. Effect of probiotic supplementation in the first 6 months of life on specific antibody responses to infant Hepatitis B vaccination. *Vaccine* 2010; 28: 2577-2579.
- Wu MY, Hill CS. Tgf-beta superfamily signaling in embryonic development and homeostasis. *Dev Cell* 2009; 16: 329-343.
- 43. Castriconi R, Cantoni C, Della CM, Vitale M, Marcenaro E, Conte R et al. Transforming growth factor beta 1 inhibits expression of NKp30 and NKG2D receptors: consequences for the NK-mediated killing of dendritic cells. *Proc Natl Acad Sci* USA 2003; 100: 4120-4125.
- Veldhoen M, Stockinger B. TGFbeta1, a "Jack of all trades": the link with pro-inflammatory IL-17-producing T cells. *Trends Immunol* 2006; 27: 358-361.
- Wan YY, Flavell RA. 'Yin-Yang' functions of transforming growth factor-beta and T regulatory cells in immune regulation. *Immunol Rev* 2007; 220: 199-213.
- Huang MC, Liao JJ, Bonasera S, Longo DL, Goetzl EJ. Nuclear factor-kappaB-dependent reversal of aging-induced alterations in T cell cytokines. *FASEB J* 2008; 22: 2142-2150.